

Effect of evening primrose oil on primeralary blood coagulation investigation in male rabbits

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الخلاصة

هدفت هذه الدراسة لمعرفة تأثير زيت زهر الربيع على معايير التخثر بعد 30 و 60 يوم من اعطاء 90 ملغم/كغم من زيت زهر الربيع لذكور الارانب السليمة. التي نتجت عن زيادة بالغة الالهمية في وقت البروثرومبين ووقت البارشيال ثرومبوبلاستين ($p < 0.01$) وعدد الصفائح الدموية قل بنسبة بالغة الالهمية ($p < 0.01$) في كلا الوقتين بينما تركيز الفايبرنوجين قل بنسبة غير مهمة. هذه التأثيرات ربما تكون نتيجة هبوط في بعض عوامل التخثر.

Abstract

This study was performed to determine the effects of Evening primrose oil (EPO) on haemostatic parameters following 30 and 60 days administration of 90 mg/kg (EPO) to healthy male rabbits. The laboratory resulting in significant increase in Prothrombin time (PT), activated partial thromboplastin time (aPTT) assays ($p < 0.01$), platelets count significantly decreased ($p < 0.01$) at both times. While fibrinogen concentration are insignificantly decreased. These effects might be due to inactivation or inhibition of factors affecting coagulation.

Introduction

Atherosclerosis is the major cause of morbidity and mortality in the developing and developed countries¹, as it is the most frequent underlying cause of coronary artery disease, carotid artery disease, and peripheral arterial disease which are resulted from superimposed thrombosis². So if thrombosis could be averted, atherosclerosis would be a much more benign disease and rarely fatal.³ Because that rupture or ulceration of an atherosclerotic plaque may precipitate the growth of platelet and fibrinaceous elements in an already narrowed lumen,⁴ growth of the fibrous plaque results in vascular remodeling, progressive luminal narrowing, blood-flow abnormalities, and compromised oxygen supply to the target organs.⁵

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Essential fatty acids are fatty acids that cannot be manufactured within the body and must be supplied by the diet.^{6,7} Evening primrose oil (EPO) provides direct and rapid supply of gamma-linolenic acid (GLA) in disease states where conversion of the dietary precursor linoleic acid to GLA is attenuated^{8,9,10} such as cardiovascular diseases¹¹ and diabetes mellitus^{12,13}. GLA (*via* DGLA which is the precursor of the prostaglandin PGH₁, which in turn forms PGE₁ and the thromboxane TXA₁ by the action of COX has anti-inflammatory, vasodilatory, and anti-aggregatory actions.¹⁴ Alternative anticoagulants which target clotting factors, like omega-3 oils from fatty fish or plant oils such as flax or canola oils have proven to be helpful in clinical trials¹⁵.

There are evidences that EPO caused remarkable improvement in clotting time; severity of atherosclerotic lesion as well significantly decreased thrombin induced platelets aggregation¹⁷. Platelets plays critical role in homeostasis, both for the formation of clot and activation of coagulation proteins¹⁸. EPO inhibits platelet aggregation in hyperlipidemic rabbits through multiple mechanisms and could be considered as antithrombotic¹⁹. These evidences show that EPO may have an effective role on haemostatic parameters hence an *in vivo* study was designed to examine the effect of EPO on prothrombin time (PT), activated partial thromboplastin time (aPTT), fibrinogen time (Fg) and platelet count.

Materials and methods

Drug treatment

EPO was obtained from vitane pharmaceutical inc., California, USA. And was taken from local pharmacy with following composition: Linoleic acid 730 mg, γ -linolenic acid 90 mg, drug were administered through oral route. The normal recommended dose of EPO is one capsule three times daily each capsule contains 1000 mg of EPO.

Animal selection

The study was carried out on twelve domestic healthy rabbits of male sex weighing from 1250-1500 grams. Animals were housed individually in cages, under controlled condition of temperature (23 \pm 2°C), and humidity (50-60%). Animals were freely access water.

Experimental design

Animals were divided in two groups, each containing six animals. a group were treated as test animals and were administered 30 mg / kg of the body weight of EPO. While controlled animals were administered water equivalent to the corresponding dose of EPO in mg/kg of the body weight. 3 ml Blood samples were collected three times, once as baseline after two week of adaptation, and the other at 30 days and last one at the end of dosing period i.e. 60 days by heart puncture of the animals.

Hematological examination:

Blood sample collection

Three ml of blood is obtained from direct heart puncture by mean of disposable plastic syringe with wide poor needle, then 1.8ml of blood but in a test tube containing 0.2ml tri-sodium citrate (3.8 %), mixture containing tube are centrifuged at 3000 rpm for 15 minute in bench centrifuge at room temperature to prepare platelet poor plasma for(pt , aptt, and fibrinogen estimation). In EDTA containing tube immediately transferring 1ml of blood (1.2 mg for 1ml blood) for platelet count.²⁰

Prothrombin time (pt)

0.1ml of prepared plasma is delivered to another glass tube placed in water bath at 37C°, and then 0.1ml of thromboplastin is added to the glass tube, then is allowed to warm for 1-3 min , then 0.1ml of pre warmed cacl₂ at 37C° is added and stopwatch is started to record end point time (clot formation).²⁰

Partial thromboplastin time (ptt)

Mixed equal volumes of phospholipid reagent and kaolin suspension were delivered in a glass tube and left in water bath 37C°. In another glass tube 0.1ml of prepared platelet poor plasma is mixed with 0.2ml of thromboplastin- kaolin solution and left for 3 min in 37C° water bath with occasional shaking. Then at exactly 3min 0.1ml of pre warmed cacl₂ is added to the mixture and stopwatch is started to record end point time (clot formation).²⁰

Platelet count

Estimation of platelet count has been carried out using Neubaur improved chamber slide and ammonium oxalate 10 mg/l as a diluents in about 1:20 ratio of dilution.²⁰

Fibrinogen Assay

Dilute plasma as 1/10 in dilution buffer. Pre warm the thrombin in water bath at 37C°. 0.2 ml of dilution incubated for 2 minutes at 37C°, then mixed with 0.2 ml of thrombin. Simultaneously start a timer and record the clotting time. Fibrinogen concentration calculated according to the calibration curve plotted on a regular graph the clotting time measured for the dilution (1/d) of tested plasma on the Y-axes. Read on the X-axis the corresponding value (a) and calculate the result as follow: Fibrinogen (mg/dl) =F*d/a (F: concentration of fibrinogen in the reference plasma ,d: reciprocal dilution of the tested plasma =10 if diluted 1/10, a: X-axis value read on calibration curve.²¹

Statistical Analysis

All data analyzed by ANOVA followed by a least significant difference (LSD).

Results

In this study there was significant increase in prothrombin time, partial thromboplastin time, in treated groups as compared with the control group, (p<0.01), as in table (1) with significantly decrease in platelets count,(p<0.01), and insignificant decrease in fibrinogen concentration in treated groups as compared with the control group, as in table (2).

Table (1) Effect of evening primrose oil on prothrombin time and activated partial thromboplastin time in compare with control group in male rabbits.

Time(day)	groups	PT in sec.	aPTT in sec.
baseline	EPO 90mg/kg	7.7500±.79183	15.717±2.0904
	control	8.1167±.79352	15.867±1.8683
30day	EPO 90mg/kg	9.9667±.78145*	24.1833±1.70695*
	control	7.9333±.79415	16.1000±1.93804
60day	EPO 90mg/kg	10.650±.7765*	25.317±1.5289*
	control	8.100±.4427	16.233±.7062

*(p<0.01) The values is Mean± Std. Deviation.

Table (2) Effect of evening primrose oil on platelets count and fibrinogen concentration in compare with control group in male rabbits.

Time(day)	groups	Platelets count (cell/l)	Fibrinogen conc. (mg/dl)
baseline	EPO 90mg/kg	230.33±19.044	214.3333 ±3.88158
	control	227.17±14.743	213.8333 ±6.61564
30day	EPO90 mg/kg	203.00±5.099*	211.16667±9.304121
	control	222.33±15.591	213.00000±3.741657
60day	EPO 90mg/kg	160.83±27.088*	205.167±6.1779
	control	218.50±14.612	214.000±8.4617

(p<0.01) * The values is Mean± Std. Deviation.

Discussion

As coagulation tests like PT, aPTT, Fg are often used to assess variation in coagulation factors²².So they can better monitor the influence of evening primrose oil on blood coagulation process. Thus we evaluate them and the effect of evening primrose oil on platelets count was also evaluated.

Prolongation in PT may be due to decrease in coagulation factors like, VII, X and V involved in extrinsic pathway. While prolongation of aPTT may be due to decrease in coagulation factors such as VIII, IX, XI, XII.²³ involve in intrinsic pathway. Present study reveals that evening primrose oil caused significant increase in PT and aPTT. Which resulted from that evening primrose oil has hypocholesterolemic effect,²⁴ and the decrease in the concentration of cholesterol may decrease the concentration of coagulation factors¹⁶.As in hypercholesterolemia, increased catabolic rate of prothrombin will stimulate hepatic synthesis of clotting factors, resulting in increased plasma concentration of clotting factors.²⁵ Also decrease absorption of lipids from gastrointestinal tract results in vitamin K deficiency,²⁶ Vitamin K deficiency cause decrease in synthesis of factors II, VII, IX and X in liver that in turn results in hypocoagulable state.^{27,28} So reduced total cholesterol by evening primrose oil may leads to vitamin K deficiency, which ultimately reduces synthesis of clotting factors resulting in prolongation of the PT. Study that asses the anticoagulant effect of EPO agree with this result and found the same prolongation in PT and PTT

also found that EPO produces its effect in a way similar to warfarin as both of them prolong the PT of the same levels. Also the study suggests that evening primrose oil may produce anticoagulant effect in the manner similar to heparin as both of them prolonged aPTT.¹⁶

Present study reveals insignificant decrease in fibrinogen level. There is evidence that increased plasma fibrinogen level has been recognized as an independent risk factor for vascular diseases.²⁹ As low thrombin concentration produces turbid fibrin clot composed of thick, loosely woven fibrin strands. While higher concentration produces fibrin clot composed of relatively thinner, more tightly packed fibrin strands.³⁰ Factor XIII (transglutaminase) increases the stability of the fibrin clot.³¹ Inflammation and platelet aggregation will increase fibrinogen production in the liver. On the other hand, Fibrinogen level increases in response to interleukins 1 and 6 (cytokines produced in arterial disorders).³² GLA has anti-inflammatory and immune-regulatory properties,^{33,34} so fibrinogen production in the liver will be decreased. Study that assesses the anticoagulant effect of EPO found that fibrinogen time are insignificantly decreased, as that decreased factor XIII concentration and thrombin concentration may affect fibrin clot structure rather its formation time.¹⁶

Present study reveals significant reduction in platelets count. There is evidence of inhibiting platelet function by evening primrose oil,³⁵ this effect may be due to GLA that stimulates PGE1 and inhibits thromboxane A2 synthesis.¹⁹ This result is disagree with other study that suggest that decrease platelet count only after 60 days with same dose (90mg/dl) and at 30 days with higher dose only, and suggest that decrease platelet count may be due to inhibition effect of evening primrose oil at initiation phase.¹⁹

Hence evening primrose oil may be decreasing coagulation factors produce this effect by inhibiting the above interaction between platelets receptors and coagulation factors V, IX and vWF resulting in platelets inhibition at their initiation phase. In response to vascular injury recruitment of platelets and interaction between platelet GPIb-V-IX and vWF takes place,³⁶ decrease platelets count by evening primrose oil may lead to decrease number of platelet receptors for thrombin, termed protease-activated receptors (PARs), since thrombin is essentially

required for activation of platelets. This suggests that thrombin-induced platelet activation is likely to be as important as platelets availability for thrombus formation in vivo.³⁶ There is a relationship between coagulation and inflammation.²⁷ Since coagulation and inflammation has been reported as biological mediators of cardiovascular disease.^{37, 38} So EPO may be of value in cardiovascular diseases, as it has anticoagulant properties that is supported by its anti-inflammatory effect, along with it's anti platelet activity.¹⁶

Conclusion

EPO has anticoagulant effect that may be as effective as other anticoagulant and anti platelets drugs that it may interact with these drugs. In addition to that it can be used as preventive measures of atherosclerosis.

References

1. Stocker R, Keaney JF (2004) Role of oxidative modifications in atherosclerosis. *Physiol Rev*, 84:1381-1478.
2. Argraves , W.S. , Tanaka ,A. , Smith ,E.P. , Twal ,W.O. Argraves , K.M. , Fan ,D. , Haudenschild , C.C (2009) : Fibulin -1 and Fibrinogen in human atherosclerotic lesions . *Histochem. Cell boil* ; 5 : 559-565.
3. Falk ,E. (2006) : Pathogenesis of atherosclerosis . *Journal of the American College of Cardiology* ; 47 (8) : C7 – C12.
4. Fuster V, Badimon L, Cohen M, Ambrose J, Badimon JJ, Chesebro JH (1988): Insights into the pathogenesis of acute ischemic syndrome. *Circulation*;77:1213-1223
5. Richard G. Lynch (2006).Milestones in Investigative Pathology The ASIP Bulletin volume 9, issue 1,16-21
6. Breanne M, Anderson and David WL Ma(2009). "Are all n-3 polyunsaturated fatty acids created equal?". *Lipids in Health and Disease* 8 (33): 33.
7. Robert S. Goodhart and Maurice E. Shils (1980) *Modern Nutrition in Health and Disease*. Lea and Febinger. Philadelphia. pp. 134-138 6th Ed.
8. Fielsend AF and Morison JIL (2000).Climatic conditions during seed growth significantly influence oil content and quality in winter and

- spring evening primrose crops (*Oenothera* spp.). *Industrial Crops and Products*, 12: 137-147.
9. Hassig A, Liang WX and Stampfli K (2000). Bronchial asthma: information on phytotherapy with essential fatty acids. *Medical Hypotheses*, 54: 72-74.
 10. Senanayake Namal SPJ and Shahidi F (2004). Incorporation of docosahexaenoic acid (DHA) into evening primrose oil (*Oenothera biennis* L.) oil via lipase-catalyzed transesterification. *Food Chemistry*, 85: 489-496
 11. Balasinska B (1998). Hypocholesterolemic effect of dietary evening primrose oil (*Oenothera paradoxa*) cake extract in rats. *Food Chemistry*, 63: 453-459.
 12. Fang C, Jiang Z and Tomlinson DR (1997). Expression of constitutive cyclooxygenases (COX.1) in rats with streptozotocin-induced diabetes; effects of treatment with evening primrose oil or an aldose reductase inhibitor on COX-1 mRNA levels. *Prostaglandins, Leukotrienes and Essential Fatty Acid*, 56: 157-163.
 13. Kuruvilla R, Peterson RG, Kincaid JC and Eichberg J (1998). Evening primrose oil treatment corrects reduced conduction velocity but not depletion of arachidonic acid in nerve from streptozotocin-induced diabetic rats. *Prostaglandins, Leukotrienes and Essential Fatty Acids*, 59: 195-202.
 14. Belch JJ, Hill A (2000). "Evening primrose oil and borage oil in rheumatologic conditions". *Am. J. Clin. Nutr.* 71 (1 Suppl): 352S-6S.
 15. Geleijnse JM, Vermeer C, Grobbee DE, (2004). "Dietary intake of menaquinone is associated with a reduced risk of coronary heart disease: the Rotterdam Study". *J Nutr.* 134 (11): 3100-5.
 16. Azra Riaz, Rafeeq Alam Khan And Shahida Parveen Ahmed. (2009), Assessment Of Anticoagulant Effect Of Evening Primrose Oil, *Pak. J. Pharm. Sci.*, Vol.22, No.4, Pp.355-359
 17. Renaud S, Gregor LM, Morazain R, Thevenon C, Benoit C, Dumont E and Mendy F (1982). Comparative beneficial effects on platelets functions and atherosclerosis of dietary linoleic and γ -linolenic acids in the rabbit, *Atherosclerosis*, 45: 43-51.
 18. Colman RW (2006). Are hemostasis and thrombosis two sides of the same coin? *J. Exp. M.*, 203: 493-495.

19. Cruz JPDL, Romero MM, Carmona JA and Villalobos MA (1997). Effect of evening primrose oil on platelet aggregation in rabbits fed an atherogenic diet. *Thrombosis Research*, 87: 141-149.
20. Dacie and Lewis (2001). Practical hematology, 9th ed., Churchill livingstone-USA;P:28,29,47,48,49,53,155,158, 206, 207, 208, 353, 354, and 355.
21. Dacie and Lewis (2006). Practical hematology, tenth ed., Churchill livingstone-USA;P:360-361
22. Yuan S, Ferrell C and Chandler WL (2007). Comparing the prothrombin time INR versus the APTT to evaluate the coagulopathy of acute trauma. *Thrombosis Research*, 120: 29-37.
23. Chan Kung-chi, Yin Mei-chin and Chao Wan-ju (2007). Effect of diallyl trisulfide-rich garlic oil on blood coagulation and plasma activity of anticoagulation factors in rats. *Food and Chemical Toxicology*, 45: 502-507.
24. Anonymous (1997), Oil of Evening Primrose. Review of Natural Products 1:1-5.
25. Miller GJ (2005). Dietary fatty acids and the haemostatic system. Review: *Atherosclerosis*, 179: 213-222.
26. Kamali F, Edwards C, Wood P, Wynne HA and Kesteven P (2001). Temporal variation in plasma vitamin K and lipid concentration and clotting factor activity in humans. *American Journal of Hematology*, 68: 159- 163.
27. Krupiczojc MA, Scotton CJ and Chambers RC (2008). Coagulation signaling following tissue injury: Focus on the role of factor Xa, *The International Journal of Biochemistry & Cell Biology*, 40: 1228-1237.
28. Shearer MJ (2009). Vitamin K deficiency bleeding(VKDB) in early infancy. *Blood Reviews*, 23(2): 49-59.
29. Ford I, Cotter MA, Cameron NE and Greaves M (2001). The effects of treatment with α -lipoic acid or evening primrose oil on vascular haemostatic and lipid risk factors, blood flow, and peripheral nerve conduction in the streptozotocin-diabetic rat. *Metabolism*, 50: 868-875
30. Wolberg AS (2007). Thrombin generation and fibrin clot structure. *Blood Reviews*, 21: 131-142.

31. Raut S, Belgrave D, Merton RE and Barrowcliffe TW (2004). Proposed 1st International Standard for Factor XIII, Plasma (02/206). Final report and recommendations. World Health Organization. Expert Committee on Biological Standardization Geneva, 15 to 19 November 2004. WHO ORGANISATION MONDIALE DE LA SANTE.
32. Zhang Z, Fuentes NL, Fuller GM. (1995) Characterization of the IL-6 responsive elements in the gamma fibrinogen gene promoter. *J Biol Chem*; 270(41): 24287-91
33. Peterson LD, Thies F and Calder PC (1999). Dosedependent effects of dietary γ -linolenic acid on rat spleen lymphocyte functions. *Prostaglandins, Leukotrienes and Essential Fatty Acids*, 61: 19-24.
34. Puri BK (2004). The clinical advantages of cold-pressed non-raffinated evening primrose oil over refined preparations. *Medical Hypotheses*, 62: 116-118.
35. Matsuo N, Osada K, Kodama T, Lim BO, Nakao A, Yamada K and Sugano M (1996). Effects of γ -linolenic acid and its positional isomer pinolenic acid on immune parameters of Brown-Norway rats. *Prostaglandins, Leukotrienes and Essential Fatty Acids*.55: 223-229.
36. Hamilton JR (2008). Protease-activated receptors as targets for antiplatelet therapy. *Blood Reviews*, 06: 1-5.
37. Rallidis LS, Georgios P, Papaioannou ML, Liakos GK, Panagiotakos DB, Anastasiadis G and Zampelas A(2004). The effect of diet enriched with α -linolenic acid on soluble cellular adhesion molecules in dyslipidaemic patients. *Atherosclerosis*, 174: 127-132.
38. Hamer M and Emmanuel S (2008). The accumulative effects of modifiable risk factors on inflammation and Haemostasis. *Brain, Behavior, and Immunity*, 22:1041-1043.