

BCG & tuberculin induced experimental lapin secondary cryoglobulinemia

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الخلاصة

جرى استخدام كل من BCG والتيوبيركلين بوصفهما منبهان مناعيان محتملان لاستثارة الاستجابة المناعية الباردة في نموذج الأرنب وتضمنت الدراسة برامج التمنيع وفرط التمنيع BCG والتيوبيركلين من خلال الاستجابة المناعية الأولية والثانوية وكانت قرائن التعرف على الاستجابة المناعية الباردة من خلال نسب الكرايوكريت في الترسيب المتعاكس بدرجة 4م والذوبان بدرجة 45 منبهان مناعيان جيدان في استثارة والاستجابة المناعية الباردة في الأرنب وظهرت زيادة الكلوبيولين البارد المحتثة تجريبيا في الأرنب ثانوية مختلطة مشتركة أو غير مشتركة مع عامل الرثوانية ويمكن تصنيفها من صنف الأرنب ثانوية على ذلك فان هذا النموذج التجريبي لزيادة الكلوبيولين البارد يوفر دعم لزيادة الكلوبيولين البارد العفوية في مرضى التدرن الرئوي .

Abstract

BCG and tuberculin were used as a possible immune stimulants for cryoresponses in a lapin model .Immunization and hyperimmunization protocols both for primary and secondary cryoimmune response were attempted .Cryocrit percentage ,protein nature through biurete reaction , ratio of cryoglobulin to total globulin of immune and hyper immune sera as well as reversible precipitable at 4c ,and solubility at 45c were identification criteria for cryoglobulin. Both BCG and tuberculin 0.05IU were found good stimulants for cryoresponse . The cryoglobulinemia was found associated and non associated with IgM anti IgG autoantibody (RF) in tuberculin case while, it was non RF associated in the case of BCG. BCG and tuberculin induce secondary mixed cryoglobulinemia of IgM-IgG-IgA type. The proposed classification was II-III class. This model stands as supportive model to the natural human mixed cryoglobulinemia associated with Pulmonary tuberculosis.

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Introduction!

Rabbits immunized with a poly valent of St. pneumoniae have produced in their sera large amount of cryoglobulins belonging to classes of IgM & IgG (1). While in a murine model. when mice injected with $1000\mu g$ of neutral K. pneumoniae polysaccharide (CPS-K).cryoglobulin was detectable in four days after injection and were most prominent after 6-10 day after injection. They were still detectable to 90 days after injection (2). Rabbits immunized with group B or group C Streptococcus show that rabbit given secondary immunization produce cryoglobulin earlier and in larger amount than rabbits given primary immunization (3). Monoclonal IgG3Rf induced cryoglobulin in vivo in murine model (4). An experimental induction of murine vasculitis induced by monoclonal antibody in which temperature sensitive immunological interaction between Fab region of cryoprotien and antigenic epitopes on the same molecules i-e an auto antibody – like reactions Wolinella succinogenes membrane **(5)**. hydrogenase in concentration of 100µg were used for immunization of mice to induce IgG3 monoclonal antibody leading to cryoglobulinemia (6). Kowalewska et al 2007have been characterizing the role of cryoglobulinemia in the development of liver disease in Thymic Stromal lymphopoietin transgenic mice that mixed cryoglobulinemia and develop hepatitis . Mice implanted with hybridoma secreting 6-19 IgG3 cryoglobulin antiIgG3 activity develop Rf with glomerulonephritis and vasculitis (7). The aim of the present work was at using tuberculin protein and BCG as possible immunostumilants for cryoglobulinemia in rabbits

Materials and methods

1- Stimulants:

The Purified Protein Derivative PPD the tuberculin ,the make of Sanofi Pasteur limited in a dilution of 0.05 IU and Bacill Calmett Gurien BCG of *Mycobacterium tuberculosis* , the make of Japan BCG laboratory in a dilution of 1/5 were used as stimulants for lapin cryoglobulinemia.

Lapin Animal Model:

Rabbits(O. caniculus) local bread brought from local market and adapted for housing condition for two weeks kept ad. Libitum of both food and drink .They were checked for and found to be free of pyrogenes and parasitis.

3-Experimental Design:

The experimental designe including:

i-Test for base- line cryoglobulin quantity in non treated and in saline! treated rabbits.

ii- Test for cryoresponse in primary immune response using single IM , single multisite Sc immunization protocol .

iii- Test for cryoresponse in secondary immune response using hyperimmunization programme of multisite injection and nasal applications (Inhalation)

4- Characterization of Cryoglobulins:

All of the scientific precusein for detection of serum cryoglobulins were in consideration as in (8). cryocrit ,cryocrit percentage ,cryoglobulin separation detection of cryoRf and determination of cryoglobulin classes were made as in(9).

Results

1- Cryoproteins:

Cryoprotein response in the sera of tuberculin immunized rabbits were found of two type. One major occurring as cryopricipitable-reverse dissolvable and one minor cryoprecipitable – no dissolvable. While these rabbits with BCG gave only the one major occurring cryoglobulins.

2- Cryoglobulin Responses:

The healthy normal Subjects were rabbit were of negative cryoglobulin responses. Like wise, two weeks adaptation with or without saline injection were of negative cryoglobulin responses. Secondary cryoresponses gave higher cryocrit percent and higher cryoglobulin concentration (Tables 1, 2). Prolonged tuberculin or BCG exposure rabbits showed high cryocrit% and higher cryoglobulin concentration (Table(1&2)) cryoglobulin/ Globulin concentration ratio were mostly one third (Table2).

3- Cryoglobulin Nature:

The onset for first appearance of cryoglobulin in lapin fresh sera were ranged between 2-5 days .The dissolving time were ranged between 1-2.30 hrs. while dissolving temperatures were ranged 37-45c (Table 1) .This both for tuberculin and BCG immunized rabbits 4-

Autoimmune and Cryoresponses:

Higher cryocrit% are associated with high IgM anti IgG autoantibody titers(Table 4). And with high IgM – IgG –IgA concentrations (Table 5) in tuberculin immunized rabbits only.

5- Immunoglobulin

Immunofixation for Cryoglobulinemia

Mixed secondary cryoglobulinemia with IgM - IgG - IgA induce by tuberculin (Table 5). It is of RF and non RF association . It can be of type II –III mixed cryoglobulinemia for tuberculin immunized rabbits . While for BCG immunized rabbits were not associated with Rf (Table 5).

Table 1: Lapin Cryoglobulin dissolving time and dissolving temperture.

Replicate			ing time IM (4)	Dissolvi multiple	Sc once		lving Time Sc & IM(8)		ving time Nasal (8)
		15 D	30 D	15 D	(4) 30 D	15 D	30 D	15 D	30 D
	D.								
1	R max	1.30	2	2	2.3	1.45	2	2.30	2.30
	Rmean	1.15	1.37	1.37	1.10	1.35	1.45	1.20	1.50
	Rmin	1	1	1	1	1.20	1.15	1.30	1.15
II	R max	2:0	2	3	2.3	2.30	2	2.30	2.15
	Rmean	12.5	1.23	2.4	1.55	1.55	1.42	1.48	1.41
	R min	1.30	1.15	2	1.30	1.30	1.15	1.20	1.30
	Replicate	Γ	Dissolving	Dissolving		DissolvingTemp.(8)		Dissolving	
	_		Temp.(5)	T	emp.(5)			i	Temp.(8)
		15 D	30 D	15 D	30 D	15 D	30 D	15 D	30 D
I	R max	45	45	45	45	45	45	45	45
	Rmean	45	45	43	45	43	45	45	45
	R min	37	45	37	45	37	45	45	45
II	R max	45	45	45	45	45	45	45	45
	Rmean	45	40.34	40.1	40.1	45	45	45	45
	R min	45	37	37	37	45	45	45	45

I = Tuberculin.

II = BCG



Table 2: The ratio of cryoglobulin to globulin in immunized and hyperimmunized rabbits.

		Cryoglobulin : Globulin!!	Ratio(%)
Single IM	Tuberculin	39 : 89	!1 : 2)
Sin	BCG	48:44	(1: 3)
ıltiple Sc	Tuberculin	19.48 : 48.24	(1: 3)
Single Multiple Sc	BCG	23.64 : 43.48	!1 : 2)
Multisite	Tuberculin	24.32 : 49.72	(1:2)
Multiple Multisite	BCG	31.88 : 55.6	!1: 2)
Nasal	Tuberculin	33.84 : 58.66	(1:2)
Ž	BCG	39.32 : 14.94	!1 : 3)

Table 3: Cryocrit % of lapin immune and hyperimmune sera with 0.05 tuberculin or 0.5 BCG.

Treatment groups		Single	IM	Single Multisite Se		Multiple Multisite IM& Se		Nasal		
		15 D	301)	15D	301)	15 D	30D	15 D	30D	!!
	R max	5.2	7	11.2	10	16	20.8	28	32	!! !!
Tuborculin	Rmcan	5	5.2	9.2	7	9.1	15.84	20.8	26.94	:: !!
Tube	Rmin	4.6	5.1	7.2	5.2	5.2	8	12	20	 !!
	R max	5.2	7.2	20.4	24	16	24	16	25.2	
	Rmean	4.7	5.8	15.90	14.36	12.56	21.36	13.6	17.96	!!
BCG	Rmin	4	4.8	10	14	9.2	16	8	16.8	!!

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Table 4: Cryoglobulin classes and Rf association in lapin model after tuberculin treatment .

Rout	Rf/titer	IgM conc.	IgG conc.	IgA conc.
Subcutaneous	+/ 16.666	2.065	8.189	2.455
		1.073	7.739	0.782
Nasal	1/ 7.714	1.603	9.377	2.279
	_	1.471	3.748	1.138

Table 5: Immunofixation of cryoglobulinemia in hyperimmunized with Tuberculin and BCG.

Groups		Ig	M	Ig	gG	Ig	IgA.	
		15 D	30 D	15 D	30 D	15 D	30 12	
M	R max	1.124	2.053	8.773	13.117	3.283	3.063	
lin Sc	Rmean	0.645	1.477	5.326	7.836	1.776	1.410	
Tuberculin Sc	R min	0.028	0.745	1.117	2.95	0.028	0.128	
Nasal	R max	2.011	4.085	13.117	14.293	2.580	3.036	
1	Rmean	0.674	1.562	5.507	7.133	2.981	1.987	
Tuberculin	R min	0.162	0.162	0.778	1.117	0.063	0.795	
N	R max	1.621	2.789	13.117	16.126	3.798	4.623	
သွ	Rmean	1.004	1.883	6.654	10.623	2.532	3.655	
BCG	R min	0.745	1.224	0.448	5.448	0.334	2.283	
Sc, IM	R max	2.011	2.011	15.506	16.126	2.564	3.798	
S S	Rmean	1.025	0.898	9.222	10.422	1.026	1.672	
BCG	R min	0.028	0.028	4.158	5.009	1.128	0.062	

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Discussion

Cryoproteins are of several types . They includes ;cryo acute phase , cryocomplement , cryofibrin , cryoglobulin and cold antibodies . Among these only cryoglobulins are of reversible perceptibility. In the present work one major reversible and one minor irreversible cryoproteins are reported (Table 1-4). The major reversible one was with following characteristics :

- i- Tuberculin or BCG induciable cryoprotein.
- ii- Secondary response gave higher cryocrit % and cryoconcentration both for tuberculin and BCG immunized rabbits .
- iii- Reversable Cryopercipitability.
- iv- Rf and non Rf associated in tuberculin while non Rf associated in BCG treated.
- v- Prolonged exposure associated with high cryo% and cryoconcentration
- vi- It is secondary mixed cryoglobulinemia of IgM IgG IgA.

Thus , it is a case of experimental mixed secondary cryoglobulinemia induced by 0.05 IU tuberculin and 1:5 BCG using different immunization protocol (Table 1-7) The proposed cryoglobuli response molecular mechanism for that RF associated is tuberculin or BCG has some epitopes are cryoglobulin active leading to monoclonal antibody including cryoglobulin in an autoantibody like mechanism (5).

While for non Rf associated cryoglobulinemia the proposed molecular mechanisms can be through thymic stromal lymphopoietin like, an interlukin 7 like cytokines with B-cells promoting properties, produce large amount of circulating cryoglobulin of mixed Igs compositions (10). Various stimulants have been tried to produce lapin cryoglobulinemia, such as *St. pneumoniae* (1), Sterptococcus group B and C (3) and *K. pneumoniae* capsular poly saccharide (2). Similarly, in murine model, the mice, bacterial membrane bond hydrogenase has been proved as stimulants for murine cryoglobulinemia (6). Others have been used monoclonal antibody IgG3 anti IgG3 Rf for induction of cryoglobulinemia(7,10). Let us put down in a comparative sense the characteristic features of human and rabbits cryoglobulin (Table 6). Thus there are basic similarities between human and rabbits cryoglobulins(

11) . Likewise there are basic similarities in the over all structures and functions of the rabbit and human immunoglobulins for classes IgM , IgG and IgA , bearing in minds the exception of subclass numbers, allotypes and idiotypes differences (12 , 13 ,14 ,15, 16,17 , 18, 19,20,21). Mean time there are marked reaction affinity between human and rabbits Rf (22 , 23,24). Thus , the determined immunoglobulin classes and RF association tables 1-8 are relative estimates and are of qualitative rather than quantitative issue . It appeared that the nature of this experimental cryoglobulinemia in the lapin model is secondary mixed with or without RF association and stayed as class II –III .Thus tuberculin 0.05IU and 1:5BCG are being reported for first time as stimulants for lapin cryoresponses.

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Table 6: Comparative view to human and rabbit cryoglobulins.

	Gebori	
Onset	2-7 Days	2-7 Days
Dissolving temperature	37c°	45c°
Reversible	+	
precipitation		
RF association	-/+	-/-
Chronicity	+	-
mixed	+	-
Secondary	+	-
Natural occurrence	ı	?
Experimental	?	-
Оссштенсе in serum	+	_
Occurence in urine	ı	?

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Table 7 classes and subclasses of human and rabbit immunoglobulin!

Species	Classes and Subclasses	
Man	IgG1-IgG2-IgG3-IgG4	IgM IgA1 –IgA2
Rabbit	IgG2 IgG1	IgM IgA1 – IgA2

From (25).

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Table 8: percent amino acid sequence homology of CH4 and Ck

!!Human	‼γ (IgG1)	α (IgA1)	‼μ(IgM)	Ск
Rabbit γ	67	30	31	-
Rabbit Cĸ				45

From Nisonoff 1982

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References

- 1. Castsoulis ,EA.; Franklin , EC. & Rothschild, M.A. 1965. Cryoglobulinemha in rabbits hyperimmunized with polyvalent pneumococcal vaccine . Immunol . 9: 327-331 .
- 2. Nakashima, I. & Kato, N. 1974 . Nobspecifiec stimulation of immunoglobulin synthesis in mice by capsular polysaccharide of *Klebsiella pneumonia* . Immunol. 27: 179-193
- 3. Herd, Z.L. 1973 .Experimental cryoglobulinemia . Immunol .25: 931-939.
- 4. Spertini, F.; Gyotoka, Y.; Shibata, T.; Izui, S. & Lambert, P. H. 1998. Experimental model of murine cryoglobulinemia induced by monoclonal antibodies and monoclonal anti- idiotype antibodies. Springer seminar. Immuno pathol. 10: 91-101.
- 5. Pastore.Y.; Lajaunins,F.; Kuroki,A.; Moll,J.; Kikuchi,S. & Izui,S. 2001. An experimental a model of cryoglobulin associated vasculitis in mice springer sen. Immunopathol .23:313-329.
- 6. Renger, J.U.; Touchard, G.; Decourt, C.; Deret, S. & Cogne, M. 2000. Heavy and light chains primary structure control IgG3 nephrogenity in an experimental model for crycrystaloglobulinlemia. Blood 95(1):3467-3472.
- 7. Kikuch, S.; Pastore. Y.; Fossati –Jimack, L.; Kuroki, A.; Yoshida, H.; Fulpius, T.; Araki, K.; Takahashi, S.; Lemoine, R.; Reininger, L. & Izui, S. 2002 .A transgenic mouse model of autoimmune glumerulonephritis and necrotizing aretritis associated with cryoglobulinemia. Immunol .169: 4644-4650.
- 8. Lynch,PLM. 2006.Audit of cryoglobulin determination in North Irlanda. Clinical biochemistry department. Althagelvin Area Hospital London derry full implementation.
- 9. Kallemuchikal, U. & Gorvic, P. 1998 . Evaluation of cryoglobulins. Arch. Pathol. Lab. Med. 123(20): 1919-1925 .

- 10. Kowalewska, I.; Muhlfeld, A.S.; Hudlkins, K.L.; Yeh, MM.; Farr, A.G.; Ravetch, J.V. & Alpers, C.E. 2007 Thymic stromal lymphopoitin transgenic mice develop cryoglobulinemia and hepatitis. Am. J. Pathol. 170: 981-989
- 11.Shnawa, I.M.S. & AL- Gebori N.R.R. 2011. Secretory and circulatory secondary cryoglobulinemia in pulmonary tuberculosis patient .J . Baby. Uni.
- 12. Nisonoff, A. 1982 Introduction to molecular Immunology . Sinauer Associate INC U.S.A.
- 13. Garcia, I.; Brandt, D.C.; Benmmar, A. & Cazenave, P-A. 1982. Basic rabbits express two types of immunoglobulins light chains κ and λ like . Proc. Nah. Acad. Sci. 79 : 4391-4394 .
- 14.Karline,S.; Ghandour,G. & Foulser,D. 1985. DNA sequence comparisons of the human,mouse & rabbit immunoglobulin kappa gene. Mol. Biol. Evol. ,2(1): 35-52.
- 15. Tizard, I. 1986. An Introduction to veterinary immunology 2nd ed.
- 16. Salzer, J.L.; Holmes, W.P. & Colman, D.R. 1987. Amino acid sequences of Myelin associated glycoprotein: Homology to the immunoglobulin gene superfamily. J. cell. Biol. 104:957 965.
- 17. Mccal, M.N. & Easterbrook Smith ,S.B. 1989. Comparison of the role of tyrosin residuces in human IgG and rabbit IgG in binding of complement C1q. Biochm. J.257: 845 851.
- 18. Bork, P.; Holm, L. & Sander, C. 1994. The immunoglobulin fold. Structural classification sequence patterns and common core. J. Mol. Bio. 242: 309-320.
- 19. Maracholonis, J.J. 1998 . Evolution of immunoglobulins Encyclopedia of Immunology v 3:1308.
- 20. Roitt, I.; Brostoff, J. & Male, D. 2001. Immunology 6th ed. Mosby. London ,221- 222.
- 21. Lefranic, C.P. ;Pommie,C. ;Ruiz,M. ;Giudicelli.V. ;Foulquier,E. ;Truong,L. ;Thouvenin-Contet,. & Lefranic .2003. IMGT unique numbering for immunoglobulin and T- cell receptor variable domains and Ig super family V like domains. Dev. Com. Immunol. 27: 55-77.

- 22.Roitt,I.M.; Hay,F.C.; Ninhan, L.J. & Male,D.K. 1982.Rheumatoid Arithritis. In Lacmann F.J.; Peter, D.K.(eds). Clinical Aspects of 23-
- 23.Fong, S.; Gilbertson, T.A. & Carson, D.A. 1983 the interal image of IgG in cross reaction anti idiotype antibodies human rheumatoid factor. J. Immunol 131:719.immunology V2 4th ed. Blackwell Scientific Publications, London 1165-1169.
- 24. Chen,pp.; Fong, S.; Houghten ,R.A. & Carson, D.A. 1985. Characterization of an epibody . J. Exp. Med .161: 323-331.
- 25. Fudenberg, H.H.; Stites, D.P.; Cladwell, J.I. & Well, J.V. 1976. Basic and clinical immunology lange. Medical publisher U.S.A. 27 30.