

The Gene expression of Combination Effects of Phage-encoded Pseudolysin Protein with Antibiotics on Gram Negative Bacterial Isolates.

Adnan Hamad Aubaid * , Hajer Majeed Abdul-Hussein **

* Microbiology Department, College of Medicine / University of Al-Qadisiyah-Iraq.

** MSc.student at Department of Microbiology, College of Medicine / University of Al-Qadisiyah-Iraq.

*Corresponding Author

Name : Adnan Hamad Aubaid

E.mail : adnan.uobeed@qu.edu.iq

Name : Hajer Majeed Abdul-Hussein

E.mail : hajermajeed18@gmail.com

Abstract

Antibiotic abuse and misuse, as well as societal and economic factors, have increased the growth of antibiotic-resistant bacteria in recent decades, rendering the medical treatment ineffective. Antimicrobial resistance kills at least 700,000 people per year around the world. Thus , the aim of present study was to evaluate the use cell wall lysis protein (endolysin) and determine if they could replace or enhancing the action of some of the current antibiotics used in treating some bacterial infections.

Different clinical specimens had been collected from patients and used to isolate and identify the bacterial isolates by using cultural , biochemical tests API systems and vitek-2 system followed by testing their resistance to antibiotics using antibiotic susceptibility test ,then estimation the gene expression of efflux pump resistance genes in three groups include tested against antibiotic alone ,tested against endolysin alone ,and against combination of antibiotic with prepared endolysin using real-time PCR.

The results of antimicrobial activity of synthetic phage encoded endolysin (1 mg/ml) against MDR *Enterobacter*-spp and *Pseudomonas*-spp showed high efficient($p<0.05$) on the expression of efflux pump genes .On the other hand the activity of endolysin along with antibiotic showed a significant($p<0.05$) decrease in the expression of efflux pump genes. The results of gene expression for efflux pump resistance genes of *Pseudomonas aeruginosa* and *Enterobacter cloacae* gave high decrease in the expression when using endolysin, and when using endolysin with antibiotic compared with control isolates

The synthetic endolysin revealed a significant effect as antimicrobial agent against MDR isolates and the effect of combination therapy of endolysin and antibiotic was synergistic effect and the expression of efflux pump genes were significantly decrease.

Keywords : gene expression; combination effects; phage-encoded pseudolysin; antibiotic; Gram-negative bacteria

Introduction

Antibiotic resistance has been referred to as “the silent tsunami facing modern medicine” (Esemu *et al.*,2022). In February 2017, WHO put some

pathogens on a list with the acronym ESKAPE (*Staphylococcus aureus*, *Acinetobacter baumannii*, *Enterococcus faecium*, *Pseudomonas aeruginosa*, *Klebsiella*

pneumonia, and *Enterobacter* species), which were given the highest "priority status" because they pose the greatest threat to humans (Mancuso *et al.*,2021).

Among the several alternative agents present currently, bacteriophage encoded peptidoglycan hydrolases, commonly known as endolysins or lysins, are a viable alternative for treating drug-resistant bacteria (Khan *et al.*,2022). Endolysins are lytic products that degrade the host bacterium peptidoglycan to release phage offspring. The exceptional capacity of endolysins to target bacterial cells when given directly is being evaluated as a potential treatment in light of antibiotic resistance (Gondil *et al.*,2020). Bacteriophages provide an appealing solution to combating rising antibiotic resistance because of their selectivity and limited range of action (Singh *et al.*,2022).

However, rather than replacing antibiotics, combining both kinds of antimicrobial agents can be more effective than using one alone. The combination strategy expected give benefit effects include enhanced bacterial suppression, better penetration into biofilms, and a limited likelihood of phage resistance (Tagliaferri *et al.*,2019). However, the choice of phage and antibiotic is experimentally established, and the influence of host variables on the efficiency is uncertain. The study looks into phage antibiotic interactions using antibiotics that have various modes of action. The findings imply that phage may reduce the working MIC for bacteria that are previously resistant to the antibiotic; however, this is dependent on the antibiotic class and pairing stoichiometry, and the host microenvironment also has a significant impact (Gu Liu *et al.*,2020).

Endolysin research is a highly dynamic area., with various possible uses being researched in the medical, veterinary, and food industries. Much of this research is being driven by the present worldwide antimicrobial

resistance challenge, with endolysins showing significant potential as antibiotic replacements or supplements. Endolysins with improved or novel features may be engineered to provide even more effective instruments (Love *et al.*,2018).

Antibiotics have enabled the treatment of formerly incurable and lethal bacterial illnesses such as meningitis and bacteremia. Antibiotic abuse and misuse, as well as societal and economic factors, have increased the growth of antibiotic-resistant bacteria in recent decades, rendering the medical treatment ineffective. Antimicrobial resistance kills at least 700,000 people per year around the world. The World Health Organization (WHO) estimates that by 2050, this figure will have risen to ten million people due to a lack of new and better therapies. Understanding these bacteria's resistance mechanisms is a crucial step in developing new antimicrobial medications to combat drug-resistant bacteria (Mancuso *et al.*,2021).

In gram-negative bacteria, the RND (Resistance Nodulation Cell Division) efflux superfamily is most commonly linked to antibiotic resistance. This efflux pump is made up of a cytoplasmic membrane-spanning transporter protein that interacts with an outer membrane-specific protein (OMP) and a periplasmic protein MFP (membrane fusion protein). Antimicrobial resistance has increased as efflux pump expression has been upregulated (AlQumaizi *et al.*,2022). The present study aimed to evaluate the combination of synthetic pseudolysin and antibiotics against multidrug resistant bacterial isolates(phenotypic and genotypic study).

Materials and Methods

Samples collection:

During the period from September 2021 To February 2022, A total of 180 patients (males &females) of different ages who admitted from different clinical

wards in Al-Diwaniyah Teaching Hospital and Burn center in Al-Diwaniyah city. The samples include swabs from Naso-gastric tube, sarak tube from intensive care unit patients, burns, oral endoscope, and foley catheter. A portable container was used to transfer all samples to the laboratory. After labeling, the swabs were streaked on MacConkey and blood agar, then incubated at 37°C for 24-48 h in an aerobic environment to perform the diagnosis, according to the standard microbiological procedure (Sharqi *et al.*,2021).

Antibiotic Susceptibility Testing

For the antibiotic susceptibility test, the antibiotic discs supplied by Bioanalyze company (Turkey). According to Clinical and Laboratory Standards Institute recommendations, the antibiotic susceptibility of the isolates was evaluated using the Kirby-Bauer disk diffusion technique (CLSI,2021). Each isolate suspension was smeared on the surface of Mueller-Hinton agar (HiMedia/India) using sterilized glass rods. The antibiotic discs were then put aseptically on the surface of the Mueller Hinton agar plate that had been inoculated. The plate was then incubated for 18 hours at 37°C. The diameters of the inhibition zones were measured according to CLSI(2021) to establish antimicrobial susceptibility. The Endolysin used antibacterial like-substance in this work was supplied from laboratory of zoonotic disease research unit / University of Al-Qadisiyah/ College of Veterinary Medicine/ Iraq.

Primers:

This work used the NCBI-Genbank database and the online primer3 plus tool for designing PCR primers. The Macrogen company in South Korea supplied these primers, which are listed in table (1)

Real-Time PCR

Table (2). shows the Real-Time PCR expression Kits that are used in the

present work & their manufacturing company and the country of origin.

Total RNA were extracted from MDR bacterial after growing in the three groups using antibiotics (*Pseudomonas aeruginosa*: ceftriaxone and *Enterobacter cloacae*: ciprofloxacin) and endolysin 1 mg (according to recommendations of previous studies) by used easy-BLUE™ Total RNA-Extraction Kit and follow the manufacturer's instructions. The total RNA that was extracted was analyzed using Nanodrop(Thermo-Scientific-NanoDrop-Lite-UV-Visible Spectrophotometer

/USA) determined the concentration of the RNA in units of ng/L and evaluated the purity of the RNA in terms of absorbance at 260/ 280 nm. The RNA that had been extracted was then treated with the DNase-I enzyme to remove any traces of genomic DNA that might have been present in the eluted total RNA. This was done with the help of samples from a DNase I enzyme kit and was carried out following the method outlined in the instructions provided by the Promega company(USA), DNase treated total RNA samples were employed in the cDNA synthesis process from mRNA transcripts by use (Accu-Power® RocketScript™ RT PreMix), this kit was performed following the instructions provided by the manufacturer Then, the RT mix components that were listed in the table above were put in the strip tubes of the Accu-Power® Rocket-Script™ RT PreMix kit. These strip tubes included all of the other components that were required for cDNA synthesis, such as primers and terminators (Reverse Transcriptase, 5X-Reaction Buffer, DTT, dNTP, and RNase Inhibitor). After that, all of the strip tubes were placed in an Exispin vortex centrifuge at 3000 rpm for 3

minutes, and then they were incubated in a Thermocycler (BioRad, USA) according to the protocol for the thermocycler conditions. The qPCR master mix was made by using a kit called RealMOD™ Green SF 2x qPCR mix Kit, which was based on the amplification of SYBER green dye in a Real-Time PCR system. After that, the components of the qPCR master mix were transferred to qPCR white plate strip tubes, exposed to an Exispin vortex, and centrifuged for a total of five minutes. Finally, they were inserted into a MiniOpticon Real-Time PCR system. The qPCR thermocycler conditions were carried out following the instructions provided in the qPCR kit, and Optimase ProtocolWriter™ was used online for primer annealing calculation purposes. The data results of qPCR were obtained for both the target genes and the housekeeping genes, and the expression analysis (fold change) was performed by using the (Δ CT methods using a reference gene) which was published by (Livak and Schmittgen, 2001), and the following equation describes the method:

$$\text{Ratio}(\text{reference} / \text{target}) = 2^{\text{CT}(\text{reference}) - \text{CT}(\text{target})}$$

Statistics analysis

The results of the current study were statistically analyzed using the Statistical Package for Social Science SPSS Twenty-Third Edition, and the statistical program was used according to the data of the study results. The Chi-square test was applied for this purpose, and significant differences less or equal to 0.05 were determined (Fortin *et al.*, 1996).

Results and Discussion

Isolation and identification of bacterial isolates

The results revealed that out of 180

isolates, only 153 (85%) gave bacterial positive growth and showed that most of them were Gram-negative bacteria which comprised a high ratio of 114 (74.50%) which included *klebsiella pneumoniae* 30 (19.60%), *Escherichia coli* 37(24.18%), *Pseudomonas aeruginosa* 19 (12.41%), *Enterobacter* spp were 20 (13.07%), *A.cintobacter bumani* 3 (1.96%) and *Citrobacter* spp 5 (3.26%).

Antibiotics susceptibility profile for isolated bacteria

Bacterial isolates were tested for their antibiotic's susceptibility toward 20 antibiotics using Kirby-Bauer disc diffusion method. Screening for antibiotic susceptibility were done according to CLSI (2021). The antibiotics which used include (Amoxiclavence, Piperacillin, Ampicillin, Imipenem, Meropenem, Cefixime, Ceftriaxone, Cefotaxime, Gentamicin, Amikacin, Ciprofloxacin, Levofloxacin, Nalidixic acid, Chloramphenicol, Tetracycline, Doxycycline, Trimethoprim, and Nitrofurantoin), and the result show that there were (47%) of *Pseudomonas* isolates and (35%) of *Enterobacter* isolates were MDR.

Relative Gene expression of resistance efflux pump genes

The reverse transcription Real-Time PCR (RT-qPCR) was done in order to molecularly detection and quantify the levels of gene expression for the efflux pump antibiotic resistance gene (*mexA* and *acrB*) in Gram negative isolates. It performed a relative expression analysis by using the Δ CT (using reference gene) method. This approach was used for the comparison of expression levels (levels of mRNA transcripts) of target genes with a housekeeping gene that was suitable. In this research it used (*rpsL* and *recA*) respectively. The analysis results of relative gene expression for efflux pump resistance genes for tested bacteria (*Pseudomonas aeruginosa* and

Enterobacter cloacae) showed the highest decrease (fold change) in the gene expression for *MexA* and *acrB*, respectively when tested with endolysin as antibacterial-like substance compared with control isolates (fig1) and (fig2).

Combination effect of Endolysin and Antibiotics

Table (3) shows the comparisons of relative gene expression of tested bacterial isolates with three treatments (endolysin 1mg ; antibiotic +endolysin ,and antibiotic alone).

This table revealed the presence of a significant decrease ($p \leq 0.05$) in transcription levels of mRNA of efflux pump gene in all tested bacterial isolates when used the combined of antibiotic and endolysin (1mg) in comparison with antibiotic alone. Also there was a significant decrease ($p \leq 0.05$) in transcription levels of mRNA of efflux pump gene when compare the used of antibiotic alone with endolysin.

The previous studies observed that endolysin and antibiotic can work together in a synergistic manner. For example, Combining colistin with LysABP-01 endolysin, which is produced as an *A. baumannii* phage, resulted in significant growth suppression and a synergistic result. Combining LysABP-01 endolysin with other antibiotics (Thummeepak *et al.*,2016). Through a minimum inhibitory concentration (MIC) and a time-kill assay, it was shown that the muralytic activity of ElyA1 endolysin in a variety of MDR strains had an elevated level of activity when used in combined with colistin. This activity was seen in a variety of *A. baumannii* and *P. aeruginosa* strains, but not in *Klebsiella pneumoniae* (Blasco *et al.*,2020). Both Gram-positive and Gram-negative bacteria were shown to benefit from the synergistic effects that antibiotics and

endolysin have on one another. Chimeric endolysin Cpl-711 was shown to have a synergic effect with a number of different antibiotics in the treatment of several drug-resistant strains of *S. pneumonia* (Letrado *et al.*,2018).

The use of endolysins as an external therapy for Gram- negative bacteria was limiting because of the presence of the outer-membrane, which blocks access to the peptidoglycan layer. In the process of creating medicines based on endolysin, one of the most significant challenges is overcoming this protective barrier. The majority causes of nosocomial infections (*Acinetobacter baumannii* and *Pseudomonas aeruginosa*). Which both are Gram-negative and have the ability to form biofilms ,and resistant to many drugs (MDR), So, certain endolysins have the ability to intrinsically pass through the outer membrane. (Love *et al.*,2018) .

Lai *et al.*(2011) recombinantly expressed LysAB2 from AB2 ,and then apply it to *A. baumannii*. An amphipathic α -helix may be found at the C-terminus of LysAB2. This helix interacts with the negatively charged components of the outer membrane, which in turn helps to facilitate the creation of a transmembrane pore. This makes it possible for the N-terminus catalytic domain to react with the peptidoglycan layer and lyse the cell, which ultimately results in antibacterial activity. The primary findings of Gu Liu *et al.*(2020) explained the following: (a) the invention of a novel high-throughput platform for rapidly assessing the effects of different phage and antibiotic concentrations on the bacterial growth, a process termed "synography" (The resulting data represent as synogram) ; (b) Synograms show a broad ranges of circumstances in which combinatorial therapy is synergistics, additive, or antagonistic, with all three occurring in same analysis at times; (c) phage may exhibit extremely efficient killing or inhibition when paired with particular

class of antibiotics ,but not with another; (e) that phage may restore antibiotic competence even with bacteria that encoded resistance elements against the antibiotic of choice, a result we call "phage adjuvation" since phage adjuvate , or make the antibiotic better; (f) that greatly genetically similar phages produce significantly different synograms even if combining them with the same group of antibiotics; (f) that phage- antibiotic synergy can avoid resistance , but just if the concentration of the antibiotic is increase ; (g) that host-like condition have a significant effect on PAS and synogram profiles in general, highlighting needing for test antibacterial effect under conditions that more reliably simulate the host environment. PAS seems to be dampened in this situation because of slower growth rate when the bacteria is in urine or blood.

The present results showed that the endolysin has a significant activity ($p \leq 0.05$) against selected Gram-negative bacterial isolates . In similar manner, the LysAB54 (100g/ml) was reported to kill logarithmic *A.baumannii* with 0.6 logs of decrease at the first minute of incubation, and more than 4 logs of reduction in the bacterial number after a 10-minute incubation (Khan *et al.*,2021). The findings revealed that Lys AB54 had both robust and quick bactericidal action, as well as strong bactericidal activity against all *A.baumannii*, *E. coli*, and *K.pneumoniae* isolates examined. Furthermore, LysAB54 susceptibility was discovered in 8 out of 10 *P. aeruginosa* strains. The variation in log reduction's antibacterial efficacy against different clinical strains might be attributed to differences in the bacterial outer membrane's molecular architecture. Notable Reductions in 4 logs (from 4.2 to 0), 2.17 logs (from 5.77 to 3.60), 2 logs (from 4.22 to 2.16), and 2.33 log (from 3.86 to 1.53) were observing in *A.baumannii* , *P.aeruginosa* , *E.coli* , and *K.pneumoniae* . These results

suggested that LysAB54 has a broad range of antibacterial activity against multi-drug resistant Gram-negative microbes.

Also Park *et al.* (2018) studied recombinant AP 50-31 and Lys B4, which displayed broad bacteriolytic activity against all the *Bacillus* spp . Oliveria *et al.*(2016), produced a recombinant endolysin (Abgp 46) from *Acinetobacter*-phage vb_Aba P_CEB1, and several multidrug-resistant *A.baumannii* strains were suppressed. They combined endolysin with a membrane-permeabilizing components to provide antibacterial action against Gram- negative bacteria including *P. aeruginosa* and *Salmonella typhimurium*.

On the other hand, Guo *et al.* (2017) observation was that Lys PA26 recombinant endolysin, expected to relate to the lysozyme-like domain family and encoded by the *Pseudomonas*-bacteriophage JD 010, demonstrated bactericidal effect against exponentially growing *P.aeruginosa* as a functions of concentration, peaking activity at 500 $\mu\text{g/mL}^{-1}$ without outer membrane-permeabilizers. *P. aeruginosa* cells were eliminated 100% at a concentration of 500 $\mu\text{g/mL}^{-1}$, compared to 20% at a concentration of 50 $\mu\text{g/mL}^{-1}$. Furthermore, after adding LysPA26 up to 50 μg , *P. aeruginosa* biofilm was dramatically decreased.

Finally, in this research an attempt was done to enhance the activity of endolysin by adding specific antibiotics to view the combination effects when using them together ,and concluded that there are a significant effect on bacterial isolate represented as change in the fold of gene expression of resistance efflux pump in each bacteria . This may help as in elimination of MDR gram negative bacteria and may also used as a treatment strategy in the future.

Conclusion

The synthetic endolysin revealed a significant effect as antimicrobial agent

against MDR isolates and the effect of combination therapy of endolysin and antibiotic was synergistic effect and the

expression of efflux pump genes were significantly decrease.

References

- bacteria. Applied microbiology and biotechnology, 90(2):529-539.
- Letrado, P.; Corsini, B.; Díez-Martínez, R.; Bustamante, N.; Yuste, J.; and García, P. (2018). Bactericidal synergism between antibiotics and phage endolysin Cpl711 to kill multidrug-resistant *pneumococcus*. Future Microbiol., 13: 1215–1223.
- Livak, K.J. and Schmittgen, T.D., 2001. Analysis of relative gene expression data using real-time quantitative PCR and the $2^{-\Delta\Delta CT}$ method. *methods*, 25(4), pp.402-408.
- Love, M.J.; Bhandari, D.; Dobson, R.C. and Billington, C., (2018). Potential for bacteriophage endolysins to supplement or replace antibiotics in food production and clinical care. *Antibiotics*, 7(1):17.
- Mancuso, G.; Midiri, A.; Gerace, E. and Biondo, C., (2021). Bacterial Antibiotic Resistance: The Most Critical Pathogens. *Pathogens*, 10(10):1310.
- Oliveira, H.; Vilas Boas, D.; Mesnage, S.; Kluskens, L.D.; Lavigne, R.; Sillankorva, S.; Secundo, F. and Azeredo, J., (2016). Structural and enzymatic characterization of ABgp46; a novel phage endolysin with broad anti-gram-negative bacterial activity. *Frontiers in microbiology*, 7:208.
- Park, S.; Jun, S.Y.; Kim, C.H.; Jung, G.M.; Son, J.S.; Jeong, S.T.; Yoon, S.J.; Lee, S.Y. and Kang, S.H., (2018). Characterisation of the antibacterial properties of the recombinant phage endolysins AP50-31 and LysB4 as potent bactericidal agents against *Bacillus anthracis*. *Scientific reports*, 8(1):1-11.
- Sharqi, H.M.; Hassan, O.M. and Obaid, A.S., (2021). Investigation of the Antibiotic-Resistant ESKAPE Pathogens in Ramadi Hospitals; Iraq. *Indian Journal of Forensic Medicine & Toxicology*, 15(4):3307.
- Singh, A.; Padmesh, S.; Dwivedi, M. and Kostova, I., (2022). How Good are Bacteriophages as an Alternative Therapy to Mitigate Biofilms of Nosocomial Infections. *Infection and Drug Resistance*, 15:503.
- Tagliaferri, T.L., Jansen, M. and Horz, H.P., 2019. Fighting pathogenic bacteria on two fronts: phages and antibiotics as combined strategy. *Frontiers in cellular and infection microbiology*, 9, p.22.
- Thummeepak, R.; Kittit, T.; Kunthalert, D.; and Sithisak, S. (2016). Enhanced antibacterial activity of *Acinetobacter baumannii* bacteriophage ØABP-01 endolysin (LysABP-01) Blasco, L.; Ambroa, A.; Trastoy, R.; Bleriot, I.; Moscoso, M.; Fernández-García, L., et al. (2020). In vitro and in vivo efficacy of combinations of colistin and different endolysins against clinical strains of multi-drug resistant pathogens. *Sci. Rep.*, 10:7163.
- Esemu, S.N.; Aka, T.K.; Kfusi, A.J.; Ndip, R.N. and Ndip, L.M., (2022). Multidrug-Resistant Bacteria and Enterobacteriaceae Count in Abattoir Wastes and Its Receiving Waters in Limbe Municipality; Cameroon: Public Health Implications. *BioMed Research International*, 2022.1:5
- Fortin, T.; Gibson, C.M.; Cannon, C.P.; Daley, W.L.; Dodge Jr, J.T.; Alexander, B.; Marble, S.J.; McCabe, C.H.; Raymond, L.; Poole, W.K. and Braunwald, E., (1996). TIMI frame count: a quantitative method of assessing coronary artery flow. *Circulation*, 93(5):879-888.
- Gondil, V.S.; Harjai, K. and Chhibber, S., (2020). Endolysins as emerging alternative therapeutic agents to counter drug-resistant infections. *International journal of antimicrobial agents*, 55(2):105844.
- Gu Liu, C., Green, S.I., Min, L., Clark, J.R., Salazar, K.C., Terwilliger, A.L., Kaplan, H.B., Trautner, B.W., Ramig, R.F. and Maresso, A.W., 2020. Phage-antibiotic synergy is driven by a unique combination of antibacterial mechanism of action and stoichiometry. *MBio*, 11(4), pp.e01462-20.
- Guo, M.; Feng, C.; Ren, J.; Zhuang, X.; Zhang, Y.; Zhu, Y.; Dong, K.; He, P.; Guo, X. and Qin, J., (2017). A novel antimicrobial endolysin; LysPA26; against *Pseudomonas aeruginosa*. *Frontiers in microbiology*, 8:293.
- Khan, F.M.; Gondil, V.S.; Li, C.; Jiang, M.; Li, J.; Yu, J.; Wei, H. and Yang, H., (2022). Bacteriophage Endolysin LysAB54 With High Antibacterial Activity Against Multiple Gram-Negative Microbes. *The Application of Phages Against Infectious Diseases*. 1:7
- Khan, F.M.; Gondil, V.S.; Li, C.; Jiang, M.; Li, J.; Yu, J.; Wei, H. and Yang, H., (2021). A novel *Acinetobacter baumannii* bacteriophage endolysin LysAB54 with high antibacterial activity against multiple Gram-negative microbes. *Frontiers in cellular and infection microbiology*, 11:70.
- Lai, M.J.; Lin, N.T.; Hu, A.; Soo, P.C.; Chen, L.K.; Chen, L.H. and Chang, K.C. (2011). Antibacterial activity of *Acinetobacter baumannii* phage φAB2 endolysin (LysAB2) against both gram-positive and gram-negative

,7:1402.

in combination with colistin. Front Microbiol

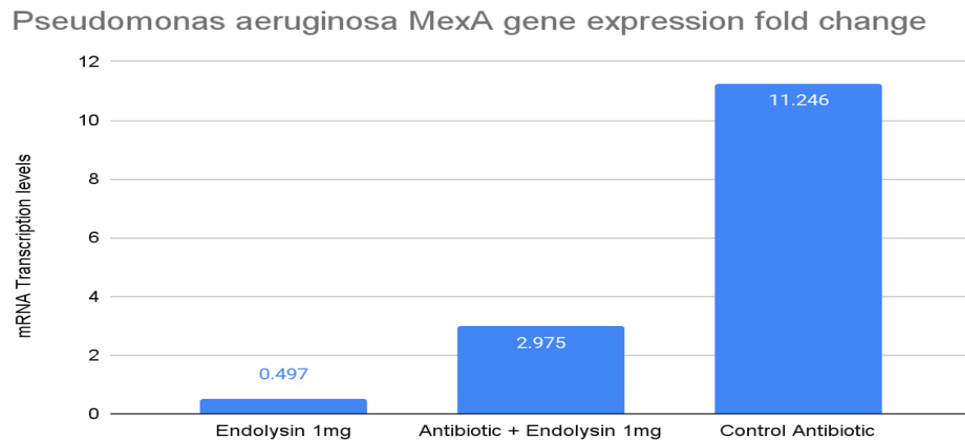


Figure (1) : Relative Gene expression of resistance efflux pump in *P.aeruginosa* under different treatment.

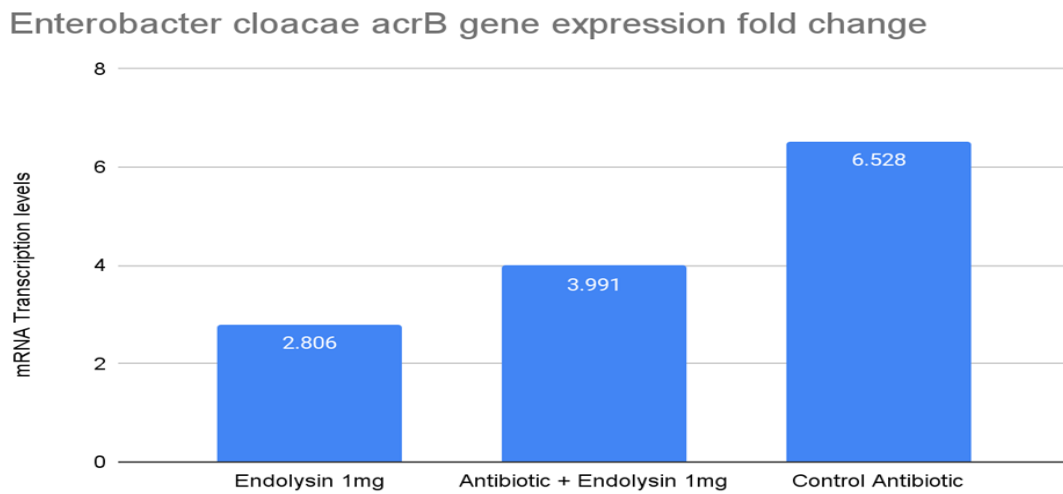


Figure (2) : Relative Gene expression of resistance efflux pump in *Enterobacter cloacae* under different treatment.

Table (1): The specific primers for target genes and their sequences.

| Primer | Sequence (5'-3') | | Product Size | Genbank |
|--|------------------|----------------------|--------------|-------------|
| <i>Enterobacter cloacae acrB</i> | F | TGTTCTGACAATGGCACAG | 133bp | DQ679966.1 |
| | R | AAGCCGTTAACCGCAAACAC | | |
| <i>Pseudomonas aeruginosa MexA</i> | F | AACAGCTCGACCCGATCTAC | 117 bp | MK341124.1 |
| | R | AGACCTTTGCCGCGTTGTC | | |
| <i>Enterobacter cloacae recA</i> <i>housekeeping gene</i> | F | GTGGTAACGCGCTGAAATTC | 111 bp | DQ679966.1 |
| | R | TGTTCTTCACGACCTTCACG | | |
| <i>Pseudomonas aeruginosa rpsL</i> <i>housekeeping gene</i> | F | TATGCACCCGCGTATACACC | 93bp | NC_002516.2 |
| | R | AAACCTCGAAACCGTTGGTC | | |

*F =Forward ; *R= Reversed

Table (2): PCR kits content used in this work with their remarks.

| PCR Kit | Companies | origin |
|--|-----------|--------------|
| easy-BLUE™ Total RNA Extraction Kit | iNtRON | South Korea |
| Trizol reagent 100ml | | |
| DNase I enzyme kit | Promega | United state |
| DNase I enzyme | | |
| 10x buffer | | |
| Free nuclease water | | |
| Stop reaction solution | | |
| Accu Power® Rocket Script™ RT PreMix | Bioneer | South Korea |
| Rocket Script Reverse Transcriptase (200U) | | |
| 5X reaction buffer | | |
| dNTP 250µM | | |
| DTT 0.25Mm | | |
| RNase Inhibitor (1U) | | |
| RealMOD™ Green SF 2X qPCR mix | iNtRON | South Korea |
| RealMOD™ Green SF 2X qPCR mix (1ml) | | |

Table (3): Gene expression of combination effect of endolysin and antibiotics.

| Bacteria | mRNA Transcription levels | | |
|-----------------------------|---------------------------|----------------------------|------------------------|
| | Endolysin 1mg | Antibiotic + Endolysin 1mg | Antibiotic As control |
| <i>P.aeruginosa</i> | 0.4±0.2 ^a | 2.9±0.8 ^b | 11.2±2.07 ^c |
| <i>Enterococcus cloacae</i> | 2.8±1.6 ^a | 3.9±1.4 ^a | 6.5±2.7 ^b |